



## SCREENING NEW SESAME (*Sesamum Indicum L.*) LINES AT TWO LEVELS OF Na Cl INCORPORATED WITH ALGAE FOR SALT TOLERANCE

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### Abstract

The development salt-tolerant sesame lines that have the resilient to the effects of climate changes are becoming an urgent need. The screening focuses on the response of 23 lines of sesame grown in a nature field and subjected to two levels of NaCl (70 and 105 mM) incorporated with algae. Experiments carry out at the Agricultural Production and Research Station, National Research Center, Egypt for two summer seasons (2017 and 2018). Six traits namely: plant height (PH), stem height to 1<sup>st</sup> capsule (SHC), fruiting zone length (FZL), number of capsules/m<sup>2</sup> (NC), seed yield/ ha<sup>-1</sup> (SYH) were used to characterize salinity tolerant. The objectives of investigate were to use algae to reduce the severe effect of salinity on sesame and to identify the lines that best adapt to salinity stress. The experiment was arranged in randomized complete blocks design with three replications for each line in both treatments. The results of combined analysis of variance indicated that interaction between lines and salinity levels was highly significant for studied traits. Mean performance of lines for studied traits were negatively affected by levels of salinity with algae compared to normal condition. While irrigation with saline water without algae the lines could not grow, which indicates that algae are important to lessen the negative effect of salinity. The correlation coefficients indicated that select index contains PH, FZL, NC and SYH could be effective to identify the salinity tolerant lines. Two lines, C8.4 and C8.8 showed the best rank and low value of stander deviation than other lines. Final produce from this study was two lines, C8.4 and C8.8 showed promising marks in studies on adaptation to saline stress, and these lines could be use as new sources of salinity tolerance in sesame. .

**Key words:** Sesame-Salinity-Algae-Correlation-Rank Correlation.

### Introduction

The crop sesame in Egypt has a great future as oil crop because possess many advantages than other oil crops, more resilience for climatic changes, drought tolerant and more income for farmers. Sesame protein has a sufficient amount of essential amino acids (methionine, cysteine and tryptophan) compared to soy (Fasuan et al.2018). Sesame oil includes sesamin and sesaminol lignans which have significant role as antioxidative activity (Xu et al. 2005 and Dar et al.2015). It also use as a cooking oil, appetizer, snack, and a flavoring agent in some foods (Myint et al.2020) .It has short season and this allows for fast cultivation, intensification, and diversification of agricultural systems (Oyeogbe et al. 2015). Sesame plays a very significant role in maintaining food and nutritional security and livelihood enhanced in developing world. But, water salinization is a major constrained to spread of sesame cultivation in Egypt. Salt aggregation suppresses plant growth and minimizes the activity to uptake water and nutrients (Hanin et al.2016). Injurious effects of Na Cl on sesame seed have been documented by several authors (Koca et al.2007, Bazrafshan and Ehsanzadeh 2014 and Bahrami et al.2016).

Plant breeding is one of the effective ways to keep crop production stable under salinity condition that posed by climatic changes. Development of new lines of oil crops such as sesame adapted to their own environmental stress conditions is important step for the sustainability development. Barhrami, et al. (2016) pointed that for a more realistic response to salt, the tolerance lines must be evaluated in ordinary field soil to facilitate the development of crops abiotic stress tolerant should be evaluated in nature filed to decrease the gab might between the knowledge gained and required. Selection and adaptation of new lines of

sesame saline stress is much significant to increase productivity, therefore, this study applied in a nature conditions.

For enhancing productivity of crop at a high level of salinity applied of exogenous growth-promoting compounds such as algae. The algae are release of bioactive compounds in algal extracts including proline, plant hormones, citric and ascorbic acid, can contribute to the mitigation of salinity stress (Ibrahim 2016). Fabio et al. (2014) announced that algae able to compromise with salinity by developed a chemical structural diversity in metabolic pathways.

Correlation coefficients provide a measure of the associations between traits and may be useful as indicators of the more important ones under considerations (Sarwar et al., 2005).To identify of salinity tolerant lines based on a single trait may be not enough; thus, rank correlation, mean rank and standard deviations of ranks for all studied traits will be used (Naghavi et al.2013). The objectives of investigate were to use algae to reduce the severe effect of salinity on 23 advanced lines of sesame and to identify the lines that best adapt to salinity stress.

### Material and Method

#### Study area

The present investigation was carried out at Agricultural Production and Research Station, Al-Nubaria district, Al-Behra Governate, National Research Center (NRC),Egypt during the two summer growing seasons ( 2017 and 2018). The nature of the soil presented in Table 1. Drip irrigation was applied weekly

**Table 1:** Physical and chemical properties of the soil of the Nubria

| Physical properties               |       |       |       |        |
|-----------------------------------|-------|-------|-------|--------|
| Soil layer depth (cm)             | 0-25  | 25-50 | 50-75 | 75-100 |
| Texture                           | Sandy | Sandy | Sandy | Sandy  |
| Course sand (%)                   | 48.66 | 55.71 | 37.76 | 37.57  |
| Fine sand (%)                     | 48.83 | 40.58 | 58.43 | 57.32  |
| Silt+ clay (%)                    | 2.51  | 3.71  | 3.81  | 5.11   |
| Bulk density (t m <sup>-3</sup> ) | 1.69  | 1.68  | 1.67  | 1.69   |
| Chemical properties               |       |       |       |        |
| EC1:5 (dS m <sup>-1</sup> )       | 0.44  | 0.53  | 1.00  | 1.56   |
| pH (1:2.5)                        | 8.60  | 8.70  | 9.32  | 9.03   |
| Total CaCO <sub>3</sub> (%)       | 7.00  | 2.34  | 4.66  | 5.02   |

**Table 2.** The origin, breeding status and description for parents

| Lines         | Breeding status | Seed source*                                | Specific characters   |
|---------------|-----------------|---|---|
| P1(HM19)      | F8-hybrid pop   | Cairo Univ.*                                | Early maturity, non branching first capsule set low, 3 capsules/axil                |
| P2 (EUL90)    | Mutant line     | Cairo Univ.*                                | Early maturity, non-branching, first capsule set low, 3 capsule/axil                |
| P3(Mutant 48) | Mutant line     | Cairo Univ.*                                | Branching, 3 capsules/axil.   |
| P4 (Giza 32)  | Local cultivar  | Ministry of Agric.& Land Reclamation, Egypt | Heavy seed weight, medium branching, one capsule/axil, long capsule, late maturity. |
| P5 (NM59)     | Exotic line     | India through IAEA**                        | Stiff stem, late maturity, one capsule/axil.  |
| P6 (Babil)    | Exotic variety  | Iraq through IAEA**                         | Low branching, 3 capsules/axil, semi- shattering capsules                           |

\* Advanced breeding materials resulted from the breeding program conducted at Agron. Dept. Fac. of Agric. Cairo Univ.

\*\* Inter. Atomic Energy Agency.

**Table 3:** Analysis of mineral analysis and amino acids of alga *Amphora coffeaeformis*

| Mineral analysis                             |      |      |      |      |         |      |      |      |                  |                               |     |
|--|------|------|------|------|---------|------|------|------|------------------|-------------------------------|-----|
| Mo   | B    | Cu   | Mn   | Zn   | Fe      | MgO  | S    | CaO  | K <sub>2</sub> O | P <sub>2</sub> O <sub>5</sub> | N   |
| 250  | 100  | 100  | 500  | 1000 | 1000    | 0.25 | 0.8  | 0.4  | 1.5              | 1.5                           | 8.0 |
| CP   | TC   | EE   | TC   | AS   | FI      | MO   |      | -    | -                | -                             | -   |
| 25.7   | 23.6 | 6.4  | 3.1  | 18.3 | 8.6     | 6.5  |      | -    | -                | -                             | -   |
| Amino acids analysis (%) on dry weight basis |      |      |      |      |         |      |      |      |                  |                               |     |
| Essential g/100g                             |      |      |      |      |         |      |      |      |                  |                               |     |
| HI   | IS   | LU   | LY   | ME   | PH      | TH   | TR   | VA   |                  | -                             | -   |
| 0.24   | 0.71 | 1.29 | 0.7  | 0.33 | 0.87    | 0.83 | 3.2  | 1.1  |                  | -                             | -   |
| Non-essential g/100g                         |      |      |      |      |         |      |      |      |                  |                               |     |
| AL   | AR   | As   | CY   | GU   | Glycine | PR   | SE   | TY   |                  | -                             | -   |
| 1.5  | 0.98 | 1.8  | 0.22 | 2.2  | 1.07    | 0.67 | 0.70 | 0.53 |                  | -                             | -   |

CP:Crude protein, TC:Total Carbohydrate,EE: Ether extract,TC: Total chlorophyll, AS:Ash, FI:Fibers, MO:Moisture Essential/100g (HI: Histidine, IS: Isoleucine,LU: Lucien, ME: Methionine, PH: Phenyl Alanine, TH:Therionine, TR:Tryptophan,VA: Valine).

Non-essential/100g (AL: Alanine, AR:Arginine, CY:Cystin, GU:glutamic, GL:Glycine,PR: Proline, SE:Serine,TY: tyrosine.

### Breeding materials and experiment design

Breeding materials used in this investigation was comprised of 23 advanced lines of sesame obtained via pedigree selection from a Agronomy Department, Faculty of Agriculture, Cairo University cooperation with Field Crops Research Department, Agricultural and Biological Division, National Research Center. Their parents' description is shown in Table 2.

The namely of lines as follows:C1.3, C1.5, C1.6,C1.8, C1.9, C1.10, C2.2, C2.3, C2.6, C3.4, C3.8,C5.7,C6.3, C6.5, C6.7, C6.9,C8.4,C8.8,C8.11,C9.6,C9.7,C9.15 and C9.20.The experiment was arranged randomized complete blocks design with three replications for each line in both treatments. The plots consisted of 1 row that was 1.5 m long and spaced 0.70

m. For the Na Cl, two concentrations were tested including 70 and 105 mM of NaCl on 23 sesame lines and one check variety (cv. Sohag) in the nature filed. Salinity levels were prepared by the put NaCl in tanks (10 m<sup>3</sup>) and used to irrigate the lines in a natural saline field. The algae were incorporated in tanks during irrigation in six equal split (at the rate 2 g/L) from germination to flowering stages. Two controls were applied, irrigation with non-saline water as control (C1) and irrigation saline water without algae as control (C2). The recommendations of Agriculture Ministry were applied.

### Algae extract specification

The locally isolated Bacillaropyhte alga *Amphora coffeaeformis* (El-Sayed et al., 2018) was massively

produced by Algal Biotechnology Unit (NRC). The obtained biomass was then dried and subjected to produce water soluble extract. Biomass was also oriented to biochemical analysis mainly ash content and amino acids. A description of the properties of algae shown in Table 3.

**Data collection:**

At harvest, 5 plants were selected in all plots and five traits were measured namely: plant height (cm), stem height to 1st capsule (cm), fruiting zone length (cm), number of capsules/m<sup>2</sup>, seed yield (t/ ha).

**Statistical analysis:**

Data were subjected to analysis of variances (Steel and Torrie 1980). Combined analysis of variance for studied traits was implemented. Duncan’s Multiple Range Test (Duncan, 1955) was applied. The coefficients of correlation between traits were estimated. These estimations were computed by computer program MSTAT-C (1991). The coefficients of correlation were tested using ‘r’ tabulated value at n-2 degrees of freedom at 5 % probability level, where n denote as number of lines studied. Rank correlation was calculated according to Spearman (1904) as follows:  $R = 1 - (\frac{6\sum D^2}{N(N^2-1)})$  Where D is rank x – rank y (i.e. the difference in the ranks) and n is the number of data pairs.

**Results and Discussion**

Combined analysis of variance for studied traits illustrated in Table 4. The salinity levels, lines and lines by salinity levels interaction variances were decomposed to provide a general formation in relation to the studied traits and mean performance of the genotypes. The salinity levels, genotypes and salinity levels × lines interaction components showed statistically highly significant variation (p < 0.05) for the studied traits. This statistical difference was due to both of the main and interaction effects of the lines with salinity levels. Moreover, Table 4 cleared that the response of the lines were unstable and fluctuated in their trait expression under the salinity levels.

The significant differences among genotypes may be due to the variations in their genetic makeup that compelled the lines to exhibits varied behavior under salinity levels. These lines need to evaluation for many years and locations due to significant of interaction between lines and salinity levels.

**Table 4:** Combined analysis of variance for studied traits at two salt concentrations.

| S.V.  | D.F | Traits    |           |          |           |             |
|-------|-----|-----------|-----------|----------|-----------|-------------|
|       |     | PH        | SHC       | FZL      | NC        | SYH         |
| Rep.  | 2   | 832.0**   | 620.6**   | 451.0**  | 729.2**   | 7952.6**    |
| A     | 1   | 66654.9** | 12788.2** | 2105.5** | 93051.0** | 1732362.6** |
| B     | 22  | 1291.3**  | 153.6**   | 787.8**  | 1465.1**  | 16777.0**   |
| AB    | 22  | 971.5**   | 289.0**   | 343.0**  | 922.3**   | 12746.0**   |
| Error | 90  | 40.4      | 17.4      | 21.0     | 30.6      | 503.0       |
| CV%   |     | 17.4      | 27.0      | 17.0     | 15.0      | 15.7        |

Mse: mean square, A: salinity levels, B: lines, AB: interaction between salinity levels and lines, Plant height (PH), Stem height to 1st capsule (SHC), Fruiting zone length (FZL), Number of capsules/m<sup>2</sup> (NB), Seed yield (t/ha) (SYH), Coefficient of variability (cv),\*\* Significant level at 0.05%.

**Quantifying the effects of salinity on sesame lines**

In this study, we applied the experiments in open field for a more realistic response to salt, the tolerance lines must be evaluated in ordinary field soil to facilitate the development of crops abiotic stress tolerant should be evaluated in nature filed to decrease the gab might between the knowledge gained and required (Barhrami, et al. 2016). Salt tolerance is described as a loop function of yield incline across a range of salt concentrations (Maas and Hoffman, 1977).

**Table 5:** Mean performance of sesame lines under two concentrations of salinity incorporated with algae for studied traits.

| 70 mM               |       |      |      |       |       |
|---------------------|-------|------|------|-------|-------|
| Genotypes           | PH    | SHC  | FZL  | NC    | SYH   |
| C1.3                | 55.0  | 19.0 | 36.0 | 110.0 | 0.219 |
| C1.5                | 45.0  | 25.0 | 21.0 | 83.0  | 0.188 |
| C1.6                | 60.0  | 25.0 | 34.0 | 111.0 | 0.208 |
| C1.8                | 58.0  | 23.0 | 33.0 | 109.0 | 0.207 |
| C1.9                | 35.0  | 15.0 | 20.0 | 61.0  | 0.158 |
| C1.10               | 63.0  | 30.0 | 32.0 | 100.0 | 0.200 |
| C2.2                | 65.0  | 29.0 | 48.0 | 130.0 | 0.225 |
| C2.3                | 55.0  | 25.0 | 30.0 | 90.0  | 0.196 |
| C2.6                | 80.0  | 25.0 | 55.0 | 140.0 | 0.230 |
| C3.4                | 52.0  | 35.0 | 17.0 | 59.0  | 0.155 |
| C3.8                | 60.0  | 30.0 | 30.0 | 105.0 | 0.201 |
| C5.7                | 30.0  | 20.0 | 10.0 | 47.0  | 0.143 |
| C6.3                | 53.0  | 26.0 | 27.0 | 88.0  | 0.191 |
| C6.5                | 55.0  | 29.0 | 26.0 | 85.0  | 0.190 |
| C6.7                | 40.0  | 30.0 | 10.0 | 70.0  | 0.161 |
| C6.9                | 69.0  | 34.0 | 35.0 | 110.0 | 0.216 |
| C8.4                | 81.0  | 15.0 | 65.0 | 180.0 | 0.250 |
| C8.8                | 61.0  | 10.0 | 50.0 | 144.0 | 0.232 |
| C8.11               | 70.0  | 22.0 | 47.0 | 133.0 | 0.222 |
| C9.6                | 50.0  | 16.0 | 31.0 | 110.0 | 0.209 |
| C9.7                | 55.0  | 27.0 | 28.0 | 90.0  | 0.194 |
| C9.15               | 69.0  | 34.0 | 35.0 | 127.0 | 0.210 |
| C9.20               | 45.0  | 27.0 | 18.0 | 60.0  | 0.156 |
| $\bar{X}$           | 56.8  | 24.8 | 32.1 | 101.8 | 0.198 |
| s                   | 50.0  | 28.0 | 22.0 | 100.0 | 121.0 |
| LSD <sub>0.05</sub> | 18.6  | 4.3  | 5.5  | 13.3  | 15.1  |
| 105 mM              |       |      |      |       |       |
| C1.3                | 43.5  | 31.0 | 14.0 | 51.0  | 100.0 |
| C1.5                | 44.0  | 33.0 | 12.0 | 39.0  | 88.0  |
| C1.6                | 52.0  | 34.0 | 20.0 | 57.0  | 99.0  |
| C1.10               | 53.0  | 44.0 | 9.0  | 40.0  | 89.0  |
| C2.2                | 56.0  | 41.0 | 16.0 | 59.0  | 105.0 |
| C3.8                | 55.5  | 40.0 | 15.0 | 60.0  | 112.0 |
| C8.4                | 69.0  | 31.0 | 38.0 | 110.0 | 138.0 |
| C8.8                | 59.0  | 31.0 | 28.0 | 100.0 | 125.0 |
| $\bar{X}$           | 54.0  | 35.6 | 18.4 | 64.5  | 105.0 |
| S                   | 0.0   | 0.0  | 0.0  | 0.0   | 0.0   |
| LSD <sub>0.05</sub> | 14.3  | 15.0 | 16.3 | 13.0  | 16.2  |
| C <sub>1</sub>      | 128.0 | 39.0 | 88.0 | 220   | 800.0 |
| C <sub>2</sub>      | 0.0   | 0.0  | 0.0  | 0.0   | 0.0   |

Plant height (PH), stem height to 1st capsule (SHC), fruiting zone length (FZL), number of capsules/m<sup>2</sup> (NB), seed yield (t/ha) (SYH), commercial cultivar(S), mean ( $\bar{X}$ ). C1: sowing in normal condition, C2: sowing under salinity conditions without algae

The results in Table 5 depicted differences among the 23sesame lines for studied traits under two levels of salinity. Studied traits were affected by the increase in salt concentration, which varied with lines (Suassuna et al.2017). Studied traits were negatively affected by two levels of salinity with algae compared to C1 While irrigation with saline water without algae (C2) the lines could not grow, indicating that algae have a negative effect on salinity.

The line C8.4 had the highest plant height under two concentrations compared S and C1 may be due to impeding normal metabolic pathways by combination of specific ion (Cl<sup>-</sup> and Na<sup>+</sup>) and osmotic and effects. Koca et al.(2007) and Bekele et al.(2017) found that salinity reduced PH. Line C8.8 showed less SHC compared to C1 and S under two concentrations. Wide range of FZL was showed under two levels of salinity whereas, five lines, C8.4, C2.6, C8.8, C8.11 and C2.2 were showed higher aeries (40cm<) than other lines, C1and S. The higher NC and SYH were recorded for two lines, C8.4 and C8.8 compared to C1 and S under two concentrations.Reduction in the NC and SYH may be due to reduction in the PH (Gaballah et al.2007 and Bahrami et al.2016).

Under 105 Mm, 8 out of 23 lines were completed their life cycle while commercial cultivar (S) did not. And the same two lines (C8.4 and C8.8) were higher, lower SHC, longer FZL, more NC and higher SYH than other lines. The reason for this may be due to these lines switch genes for salinity tolerant in the presence of algae. Islam et al. (2016)

### Effects of algae on salinity level

Algae extract found that able to overcoming the adverse effects of saline stress on plants, possibly due to the presence of proline in algae (Table 3). Algae help the plant tolerate the stress (Pazuki et al.2015 and El-Sayed et al. 2018).Algae were early considered as an important group of microorganisms capable of fixing atmospheric nitrogen in addition enhance the tolerance to stress conditions (Ghalab and Salem, 2001). Algae cause significant increase in root growth, fresh and dry weights of roots, total biomass, yield component, photosynthetic pigments and growth promoting hormones (Abdel-Maguied et al., 2004 and Enan et al.2016). Algae extract was considered as a source of nutrition for sustainable agriculture especially in the newly reclaimed soil. Chemical analysis of algae extract revealed the presence of a wide variety of plant growth regulators such as auxins and cytokinins (Zhang and Ervin, 2004) which in turn stimulate root establishment, root elongation and promoting vegetative growth of plants. In addition, algae extract induce many positive effects in concern improve crop yield and quality, increase nutrient uptake, resistance to frost and stress conditions. As a result of the functional activity, there is an increase of photosynthetic apparatus through raising the contents of total carbohydrates, starch, amino acids and protein (Raupp and Oltmanns, 2006 and Yassen et al., 2007). In this study we found that algae have a negative effect on the high salinity level in sesame production. Thus, sesame can be grown in areas affected by salinity.

### Correlation analysis

To determine traits more related to seed yield under stress, the correlations between studied traits were calculated. Sserumaga et al. (2018) pointed that direct selection for high seed yield in normal condition may be leads to improve seed yield in stress condition. Data in Table 6 showed that PH had positive and highly significant correlation with FZL, NC and SYH.

FZL was positively correlated with NC and SYH while non significant negative correlation was found between SHC and FZL. As expect, the highest positively correlated recorded between NC and SYH because NC is the main component of sesame seed yield components. The

pointed that exposure to salt and biotic factors were decreased sesame seed yield ha<sup>-1</sup> around 300–400 kg/ha. Aghajari et al.(2014) found that grains being decreased by 66% when plants were irrigated with water of 55 mM of Na Cl compared to normal condition. Mahmood et al. (2003) pointed that seed yield decreased under high salinity level caused by the abscisic acid, because ABA synthesis in plants under saline stress, as a result, the flowers and fruits fall down. Reduction in the seed yield/ha-1 under salinity conditions may be due to many reasons: it could be due to drought stress caused by the salt effect on water system in plant tissues and cell magnification and increase in leaf senescence and abscission (Jaleel et al. 2008),ionic imbalances (Khan et al.2000), reduction in protein synthesis and chlorophyll content (Srivastava et al.2008).As a result, decrease in cell division,wilting ,closure of stomata and decrease of water content in the cell (Tester and Davenport 2003). From this study, two lines (C8.4 and C8.8) showed a good performance for studied traits under salinity levels compared to check variety, so it could be use as new sources of salinity tolerance in sesame.

significant of correlation coefficients under two concentrations were same. In short, the select index that contains traits, PH, FZL, NC and SYH will be effective to identify the salinity tolerant lines of sesame.

### Ranking of lines for salt tolerance

To identify of salinity tolerant lines of sesame based on a single trait may be not enough therefore, rank, mean rank and standard deviations of ranks for five traits were calculated (Table 7).

The results in Table 7 indicated that two lines, C8.4 and C8.8 recorded the best rank and low SD than other lines under two concentrations, consequently classified as the most salinity tolerant lines. Bekele et al.(2017) ranked accessions of Ethiopian sesame for salt tolerance based on nine traits while both Farshadfar et al., (2012) and Naghavi et al., (2013) were used the rank correlation to identify of drought tolerant in wheat and maize.

Table 6: Simple correlation between studied traits at two concentrations of salinity

| Traits | SHC  | FZL     | NC     | SYH         |
|--------|------|---------|--------|-------------|
|        |      | 70 mM   |        |             |
| PH     | 0.27 | 0.85**  | 0.62** | 0.60**      |
| SHC    |      | -0.26   | 0.21   | 25.0        |
| FZL    |      |         | 0.51** | 0.47**      |
| NC     |      |         |        | 0.93**      |
|        |      | 105 mM  |        |             |
| PH     | 0.10 | 0.81**  | 0.84** | 0.85**      |
| SHC    |      | -0.59** | 0.25   | 0.27        |
| FZL    |      |         | 0.96   | 0.92        |
| NC     |      |         |        | <b>0.97</b> |

Plant height (PH), stem height to 1st capsule (SHC), fruiting zone length FZL), number of capsules/m<sup>2</sup> (NB), seed yield (t/ha) (SYH), \*\* P <0.05

**Table 7:** Rank, ranks mean (R) and standard deviation of ranks (SD) of new sesame lines for studied traits under 70 and 105 mM of NaCl incorporated with algae in nature field.

| Genotypes | PH   | SHC  | FZL   | Traits |      |      |     |
|-----------|------|------|-------|--------|------|------|-----|
|           |      |      |       | NC     | SYH  | R    | SD  |
|           |      |      | 70mM  |        |      |      |     |
| C1.3      | 13.0 | 5.0  | 19.0  | 11.0   | 7.0  | 11.0 | 5.5 |
| C1.5      | 21.0 | 10.0 | 18.0  | 19.0   | 18.0 | 17.2 | 4.2 |
| C1.6      | 8.0  | 11.0 | 9.0   | 7.0    | 9.0  | 8.8  | 1.5 |
| C1.8      | 10.0 | 9.0  | 10.0  | 13.0   | 10.0 | 10.4 | 1.5 |
| C1.9      | 23.0 | 3.0  | 20.0  | 21.0   | 21.0 | 17.6 | 8.2 |
| C1.10     | 7.0  | 9.0  | 7.0   | 14.0   | 13.0 | 10.0 | 3.3 |
| C2.2      | 6.0  | 8.0  | 5.0   | 4.0    | 3.0  | 5.2  | 1.9 |
| C2.3      | 14.0 | 12.0 | 12.0  | 15.0   | 15.0 | 13.6 | 1.5 |
| C2.6      | 2.0  | 13.0 | 2.0   | 5.0    | 2.0  | 4.8  | 4.8 |
| C3.4      | 12.0 | 23.0 | 22.0  | 23.0   | 22.0 | 20.4 | 4.7 |
| C3.8      | 9.0  | 19.0 | 13.0  | 12.0   | 11.0 | 12.8 | 3.8 |
| C5.7      | 24.0 | 6.0  | 23.0  | 24.0   | 23.0 | 20.0 | 7.8 |
| C6.3      | 17.0 | 14.0 | 15.0  | 17.0   | 17.0 | 16.0 | 1.4 |
| C6.5      | 15.0 | 18.0 | 16.0  | 18.0   | 19.0 | 17.2 | 1.6 |
| C6.7      | 19.0 | 15.0 | 24.0  | 20.0   | 8.0  | 17.2 | 6.1 |
| C6.9      | 4.0  | 20.0 | 7.0   | 8.0    | 14.0 | 10.6 | 6.4 |
| C8.4      | 1.0  | 2.0  | 1.0   | 1.0    | 1.0  | 1.2  | 0.4 |
| C8.8      | 8.0  | 1.0  | 6.0   | 2.0    | 4.0  | 4.2  | 2.9 |
| C8.11     | 3.0  | 7.0  | 4.0   | 3.0    | 12.0 | 5.8  | 3.8 |
| C9.6      | 17.0 | 4.0  | 11.0  | 9.0    | 16.0 | 11.4 | 5.3 |
| C9.7      | 11.0 | 17.0 | 14.0  | 16.0   | 5.0  | 12.6 | 4.8 |
| C9.15     | 5.0  | 22.0 | 8.0   | 6.0    | 20.0 | 12.2 | 8.1 |
| C9.20     | 22.0 | 16.0 | 21.0  | 22.0   | 24.0 | 21.0 | 3.0 |
| S         | 18.0 | 16.0 | 17.0  | 10.0   | 6.0  | 13.4 | 5.2 |
|           |      |      | 105mM |        |      |      |     |
| C1.3      | 8.0  | 2.0  | 6.0   | 6.0    | 4.0  | 5.0  | 2.6 |
| C1.5      | 7.0  | 4.0  | 7.0   | 8.0    | 8.0  | 6.4  | 2.5 |
| C1.6      | 6.0  | 5.0  | 3.0   | 5.0    | 5.0  | 4.4  | 1.3 |
| C1.10     | 5.0  | 8.0  | 8.0   | 7.0    | 7.0  | 7.0  | 1.2 |
| C2.2      | 3.0  | 7.0  | 4.0   | 4.0    | 4.0  | 4.0  | 0.7 |
| C3.8      | 4.0  | 6.0  | 5.0   | 3.0    | 3.0  | 4.2  | 1.3 |
| C8.4      | 1.0  | 3.0  | 1.0   | 1.0    | 1.0  | 1.2  | 0.4 |
| C8.8      | 2.0  | 1.0  | 2.0   | 2.0    | 2.0  | 2.2  | 0.4 |

Plant height (PH), stem height to 1st capsule (SHC), fruiting zone length (FZL), number of capsules/m<sup>2</sup> (NB), seed yield (t/ha) (SYH), S:commercial cultivar, R: ranks mean, SD: standard deviation of ranks.

## Conclusion

In this study, 23 advanced lines of sesame were screened in a nature field at two levels of Na Cl (70 and 105 mM ) with add algae for five traits. We found that sesame lines respond differently to salt stress. Five traits were negatively affected by salinity levels with add algae. But at high level of salinity without algae the lines did not grow indicated that the significant role of algae on salinity stress may be due to release of bioactive compounds which contributed to the mitigation of salinity stress or contributed to catalyzed genes in lines. Based on overall and ranks of studied traits two lines, C8.4 and C8.8 were promising for studies on adaptation to saline stress in sesame.

## References

- Abdel-Maguid AA, El-Sayed AB, Hassan HS. 2004. Growth enhancement of olive transplants by broken cells of fresh green algae as soil application. *Minufiya J Agric Res* 29: 723-733.
- Aghajari, S., Boroomand-Nasab, S., Sakinejad, T., Behmanesh, M. and Motamedi, B. 2014. Sesame (*Sesamum indicum L.*) performance under different salinity levels of water. *Researcher*. 6, Abdel-Maguid, A. A.; El-Sayed, A.B. and Hassan, H. S. (2004).
- Growth enhancement of olive transplants by broken cells of fresh green algae as soil application. *Minufiya J. Agric. Res.*, 29 (3) 723-73321-24.
- Bahrami, H., A. O. Jafari, J. Razmjoo.2016. Effect of salinity levels (NaCl) on yield, yield components and quality content of sesame (*Sesamum indicum L.*) cultivars. *Envir. Manag. and Susta. Devel.*5(2):104-117.
- Bazrafshan, A.H. and P. Ehsanzadeh.2014. Growth, photosynthesis and ion balance of sesame (*Sesamum indicum L.*) genotypes in response to NaCl concentration in hydroponic solutions. *Photosynthetica*. 52: 134–147.
- Bekele,A., Besufekad, Y., Adugna S., Yinur, D.2017. Screening of selected accessions of Ethiopian sesame (*Sesame indicum L.*) for salt tolerance. *Biocata. and Agric. Biotec.*9:82-94.
- Dar, A.A., Verma N.K.and Arumugam N.2015. An updated method for isolation, purification and characterization of clinically important antioxidant lignans—sesamin and sesamol from sesame oil. *Ind Crop Prod.* 2015;64:201–208
- Duncan, D.B. 1955. Multiple range and multiple tests. *Biometrics*. 11:1-42.
- El-Sayed AB, Shehata SA, Taha, Sahar S, Hamouda HA, Abdelgawad Karima F, Youssef Doaa M (2018) Algae extract overcoming the adverse effects of saline stress

- in hydroponic grown tomato plants. *J Food Agric Environ* 16(2):92–99.
- Enan, S.A.A.M., A. M. El-Saady and A. B. El-Sayed .2016.Impact of Foliar Feeding With Alga Extract and Boron on Yield and Quality of Sugar Beet Grown in Sandy Soil. *Egypt. J. Agron.*38(2): pp.319-336.
- Fabio, A.E., B.C. Torresa, G. Thais, B.C. Passalacqua, M.A. Angela, B.C. Velasqueza, A. Rodrigo, D. Souza, P. Colepicolod, A.S. Márcia, C. Graminha. 2014. New drugs with antiprotozoal activity from marine algae: a review. *Rev Bras Farmacogn.* 24: 265-276.
- Farshadfar, E., B. Jamshidi and M. Aghaee, 2012. Biplot analysis of drought tolerance indicators in bread wheat landraces of Iran. *Inte. J. of Agric. and Crop Sci.*, 4, 226-233.
- Fasuan, O.T., Taiwo O. Omobuwajo, Saka O. Gbadamosi .2018.Characterization of protein isolate from *Sesamum indicum* seed: In vitro protein digestibility, amino acid profile, and some functional properties. *Food Sci Nutr.*6:1715–1723.
- Gaballah, M. S., B. Abu Leila, H. A. El-Zeiny and S. Khalil.2007. Estimating the performance of salt-stressed sesame plant treated with antitranspirant. *J. of Applied Sci. Res.*9: 811-817.
- Ghalab AM, Salem SA (2001) Effect of bio-fertilizer treatments on growth, chemical composition and productivity of wheat grown under different levels of NPK fertilization. *Ann Agric Sci* 46:485–509.
- Hanin, M., C. Ebel, M. Ngom, L. Laplaze and K.Masmoudi.2016. New insights on plant salt tolerance mechanisms and their potential use for breeding. *Fron. in Plant Sci.*7:1-17.
- Ibrahim M. W. 2016. Potential impact of marine algal extracts on the growth and metabolic activities of salinity stressed wheat seedlings. *J. of Appl. Sci.* 16 (8): 388-394.
- Islam, F.; R.A.Gill.; B. Ali.; M.A. Farooq.; L. Xu.; U. Najeeb.; W. Zhou.2016. Sesame. In: *Breeding oilseed crop for sustainable production: opportunities and constraints*; Gupta, S.K., Ed.; Academic Press: Cambridge, MA, USA, pp. 135–147.
- Jaleel, C. A., Gopi, R., Manivannan, P., & Panneveerselvam, R. (2008). Soil salinity alters the morphology in *carthamus roseus* and its effects on endogenous mineral constituents. *EurAsia. Journal of Biosciences*, 2, 18-25.
- Khan, M. A., Ungar, I. A., Showalter, A. M. (2000). Effect of salinity on growth, water relations and ion accumulation of the subtropical perennial halophyte, *Atriplex griffithii* var *stocksii*. *Annals of Botany*, 85, 225-232. <http://dx.doi.org/10.1006/anbo.1999.1022>
- Koca, H., M. Bor, F.Ozdemir, I. Turkan.2007. The effect of salt stress on lipid peroxidation, antioxidative enzymes and proline content of sesame cultivars. *Environ. Exp. Bot.* 60: 344–351.
- Maas, E. V., and Hoffman, G. J. (1977). Crop salt tolerance-Current assessment. *J. Irrig. Drainage Div. ASCE* 103(IR2), 115-134.
- Mahmood, S., Iran, S. and Athar, H.R. 2003. Intra-Specific Variability in Sesame (*Sesamum indicum* L.) for Various Quantitative and Qualitative Attributes under Differential Salt Regimes. *J. of Res. Sci.* 14:177-186.
- MSTAT-C program, 1991. A software program for the design, management and analysis of Agronomic research experiments. Michigan State University.
- Myint, D., Syed A. Gilani, Makoto Kawase and Kazuo N. Watanabe.2020. Sustainable Sesame (*Sesamum indicum* L.) Production through Improved Technology: An Overview of Production, Challenges, and Opportunities in Myanmar. *Sustainability.*12:1-21.
- Naghavi, M.R., A. P. Aboughadareh and M. Khalili, 2013. Evaluation of drought tolerance indices for screening some of corn (*Zea mays* L.) cultivars under environmental conditions. *Not. Sci. Biol.*, 5(3):388-393
- Oyeogbe, A., R. Ogunshakin, V. S. Epatel, B. Towards.2015. Sustainable intensification of sesame-based cropping systems diversification in northwestern India. *J. of Food Secu.* 3: 1-5.
- Pazuki, A; Asghari, J; Sohani, M; Pesarakli, M & Aflaki, F (2015). "Effects of Some Organic Nitrogen Sources and Antibiotics on Callus Growth of Indica Rice Cultivars". *Journal of Plant Nutrition.* 38 (8): 1231–1240. doi:10.1080/01904167.2014.983118.
- Raupp, J. and Oltmanns, M. (2006) Farmyard manure, plant based organic fertilizers, inorganic fertilizer-which sustains soil organic matter best. *Aspects of Applied Biology*, 79, 273-276.
- Sarwar, G.; Haq M. A. and Mughal, M. S. (2005). Genetic parameters and correlation study in diverse type of sesame Germplasm. *Sesame and safflower Newsletter* 20 : 1-4.
- Spearman C.E., 1904. The proof and measurement of association between two things. *Ame. J. of Psy.*,15: 72–101.
- Srivastava, A. K., Bahargava, P., Thapar, R., Rai, L. C. (2008). Salinity-induced physiological and proteomic changes in *Anabaena doliolum*. *Environmental and Experimental Botany*, 64, 49-57. <http://dx.doi.org/10.1016/j.envexpbot.2007.12.012>
- Sserumaga, J.P., Y. Beyene, K. Pillay, A. Kullaya, S. O. Oikeh, S. Mugo, L. Machida, I. Ngolinda, G. Asea, J. Ringo, M. Otim, G. Abalo and B. Kiula, 2018. Grain-yield stability among tropical maize hybrids derived from doubled-haploid inbred lines under random drought stress and optimum moisture conditions. *Crop and Past. Sci.*, 69(7): 691-702.
- Steel RGD, Torrie JH. 1980. Principles and Procedures of Statistics. 2nd ed. McGraw Hill Book Co. Inc., New York.
- Suassuna, J. F., F. P. Dantas, B. M. E. Barbosa, A. N. H., C., M. A. Soares de, J. D. Fernandes .2017. Tolerance to salinity of sesame genotypes in different phenological stages. *Amer. J. of Plant Sci.* 8: 1904-1920.
- Tester, M., & Davenport, R. (2003). Na<sup>+</sup> tolerance and Na<sup>+</sup> transport in higher plants. *Annals of Botany*, 91, 503-527. <http://dx.doi.org/10.1093/aob/mcg058>.
- Xu, J., S. Chen and Q. Hu .2005. Antioxidant activity of brown pigment and extracts from black sesame (*Sesamum indicum* L.) seed. *Food Chems.*91:79-83.
- Yassen AA, Badran NM, Zaghoul SM (2007) Role of some organic residues as tools for reducing metals hazard in plant. *World J Agric Sci* 3(2):204–209
- Zhang, X. and Ervin, H. (2004) Seaweed extract and humic acid contain cytokinins. *Crop. Sci.* 44 (5), 1509.